OTES

NPS-indole derivatives in most solvents. After chromatography of this treated ixture in solvent D the  $R_F$  values for the four DNPS-derivatives were found identical ) standard compounds either alone or in mixture.

We thank Mr. G. K. McCully for valuable help with the chromatographic eparations.

Itlantic Regional Laboratory, Iational Research Council of Canada, Ialifax, Nova Scotia (Canada)

R. K. RAI\* O. HUTZINGER\*\*

I R. K. RAJ AND O. HUTZINGER, in preparation.

2 A. FONTANA, F. MARCHIORI, R. ROCCHI AND P. PAJETTA, Gazz. Chim. Ital., 96 (1966) 1301.

3 E. SCOFFONE, A. FONTANA AND R. ROCCHI, Biochemistry, 7 (1968) 971.

4 K. PODUŠKA, Collection Czech. Chem. Commun., 33 (1968) 3779. 5 G. W. PEROLD AND H. L. F. SNYMAN, J. Am. Chem. Soc., 73 (1951) 2379. 6 C. BROWN AND D. R. HOGG, J. Chem. Soc., (B), (1968) 1315.

Received July 7th, 1969

\* Royal Society Nuffield Foundation Commonwealth Fellow and National Research Council of Canada Visiting Scientist. Permanent address: Division of Biochemistry, University of Kerala, Trivandrum, India.

\*\* Please address reprint requests to this author.

J. Chromalog., 44 (1969) 199-201

### CHROM. 4283

## Polyamide-silica gel thin-layer chromatography of food preservatives

The chromatography of food preservatives has been studied by numerous investigators. The separation of these preservatives on thin layers of cellulose acetatepolyamide<sup>1</sup>, cellulose<sup>2</sup> and silica gel<sup>3</sup> has been reported, but there is no report on separation by polyamide-silica-gel layers. In a previous report<sup>4</sup>, better separation of red food dyes was obtained with polyamide-silica gel layers; therefore, this method was further applied to separate ten preservatives. For comparison, the thin-layer chromatography of only polyamide and of only silica gel is also described.

### Experimental

Preparation of polyamide-silica gel mixed layer. Ten grams of polyamide ( $\varepsilon$ -polycaprolactam CM 1007S of Toyo Rayon Co., Tokyo, Japan) were dissolved in 80 ml of 90% formic acid; then 20 ml of distilled water were added. After gentle warming (below 40°) and stirring, a homogeneous solution was obtained. It was then cooled to room temperature, and 52 g of Silica Gel G (E. Merck) were added. Of the previous solution 200 ml were poured into a dish  $(14.5 \times 19.5 \times 2.5 \text{ cm})$  into which a glass plate  $(12 \times 14 \times 0.1 \text{ cm})$  was dipped. Both sides of the glass were covered homogeneously. The glass was hung for 2 min over the dish to let the excess solution drain off. It was then air dried for 3 h and heated at  $100^{\circ}$  for 30 min. These layers can be stored for a long period.

*Preparation of polyamide layer*. Instead of 10, 20 g of polyamide were dissolved before proceeding as described in the previous method, but without adding Silica Gel G.

Preparation of silica-gel layer. Dilute slurries of Silica Gel G (45 g to 100 ml of water) were sprayed at 2 kg/cm<sup>3</sup> pressure from a distance of 20 cm onto 8 horizontal glass plates ( $12 \times 14$  cm) which were then dried at 100° for 30 min. The thickness of the layers was about 250  $\mu$ .

Chromatographic procedure. A 0.2% alcoholic solution of samples was applied to the starting line 1.5 cm from the bottom of the layer, and the plate was developed by ascending techniques. The chamber had been equilibrated with the respective solvent for 30 min before use.

*Visualization.* The layers were sprayed with a 0.07% alcoholic solution of Rhodamine B, and deep-violet spots could be observed under UV light at 366 m $\mu$ .

### Results and discussion

 $R_F$  values obtained with two solvent systems are given in Table I. It is interesting to note that the  $R_F$  values of the *p*-hydroxybenzoic acid ester in the two solvent systems are reversed. In the nonaqueous system (solvent I), the  $R_F$  values increase with an increase in the molecular weight of the esters, *i.e.* the order of  $R_F$  values is the same as that obtained by previous workers<sup>1-3</sup>. However, in the aqueous system (solvent II), the  $R_F$  values decrease with an increase in the molecular weight.

In both solvent systems, when using polyamide-silica gel mixed layers the  $R_F$  values are lower and separation is better than when polyamide and silica gel layers are employed. A 10-cm ascent from the origin is more rapid using the mixed layers

### TABLE I

#### CHROMATOGRAPHIC DATA

Solvent I: *n*-hexane-benzene-glacial acetic acid (1:1:1); solvent II: water-28% ammonia solution (20:5). a,  $R_F$  value obtained on polyamide-silica gel layer; b, silica gel layer; c, polyamide layer.

No.	Substance	Solvent I			II		
		a	ь	c	a	ь	c
I	Methyl p-hydroxybenzoate	0.30	0.73	0.61	0.55	0.91	0.78
2	Ethyl p-hydroxybenzoate	0.35	0.77	0.69	0.46	0.88	0.70
3	Propyl p-hydroxybenzoate	0.41	0.78	0.74	0.34	0.79	0.59
4	Isopropyl p-hydroxybenzoate	0.44	0.78	0.74	0.32	0.77	0.60
5	Butyl p-hydroxybenzoate	0.49	0.79	0.78	0.24	0.69	0.49
6	Isobutyl p-hydroxylbenzoate	0.53	0,80	0.81	0.21	0.69	0.50
7	Sorbic acid	0.88	0.87	0.98	0.73	0.95	0.94
8	Benzoic acid	0.84	0.89	0.98	0.64	0.96	0.88
9	Salicylic acid	0.66	0.84	0.62	0.49	0.96	0.74
10	Dehydroacetic acid	0.74	0.96	0.80	0.62	0.95	0.85
Time required (min) <sup>a</sup>		90	25	240	130	15	180

<sup>a</sup> Time required to ascend 10 cm from origin.

#### OTES

an using polyamide layers. The content (16%) of polyamide in this polyamide-silica el mixed layer is higher than that in the previous report  $(12\%)^4$  for getting a more able layer.

# epartment of Pharmacy, Taipei Medical College, aipei, Taiwan (Republic of China)

HUNG-CHEH CHIANG

T. SALO AND K. SALMINEN, Z. Lebensm.-Untersuch.-Forsch., 124 (1964) 448.

2 J. W. COPIUS-PEEREBOOM AND H. W. BEEKES, J. Chromatog., 14 (1964) 417.

3 H. GANSHIRT AND K. MORIANZ, Arch. Pharm., 293 (1960) 1065.

H.-C. CHIANG, J. Chromatog., 40 (1969) 189.

irst received May 12th, 1969; revised manuscript received July 1st, 1969

J. Chromatog., 44 (1969) 201–203

## HROM. 4284

## olyamide-kieselguhr thin-layer chromatography of yellow food dyes

The thin-layer chromatography of food dyes has been studied by numerous nvestigators. The separation of synthetic food dyes by thin layers of cellulose<sup>1</sup>, tarch<sup>2</sup>, silica gel<sup>3</sup>, aluminum oxide<sup>4</sup> and polyamide<sup>5</sup> has been reported. Recently, a etter separation of red food dyes was obtained by CHIANG<sup>6</sup> with a mixed polyamideilica gel thin layer. Therefore, further modification of this method was tried. In this ote, the separation of five yellow food dyes and three harmful yellow dyes (auramine, netanil yellow and picric acid) by mixed polyamide-kieselguhr thin-layer chromatoraphy is described. For comparison, the thin-layer chromatography on only polymide and on only kieselguhr is also described.

### Experimental

Preparation of polyamide-kieselguhr mixed layer. Ten grams of polyamide chip Nylon 6, type 1022B of UBE Industrial Ltd., Osaka, Japan) were dissolved in 80 ml f 90% formic acid; then 20 ml of distilled water were added. After warming (below 0°) and stirring, a homogeneous solution was obtained. It was then cooled to room emperature, and 40 grams of Kieselguhr G (E. Merck) were added. Of the previous olution 200 ml were poured into a dish  $(14.5 \times 19.5 \times 2.5 \text{ cm})$  into which a glass plate  $12 \times 14 \times 0.1 \text{ cm}$ ) was dipped. Both sides of the glass were covered homogeneously. The glass was hung for 2 min over the dish to let the excess solution drain off. It was hen air dried for 3 h and heated at 100° for 30 min.

Preparation of polyamide layer. Instead of 10 g, 20 g of polyamide were dissolved before proceeding as described in the previous method, but without adding Kieselguhr G.

Preparation of kieselguhr layer. Dilute slurries of Kieselguhr G (45 g in 100 ml of water) were sprayed at 2 kg/cm<sup>3</sup> pressure from a distance of 20 cm onto 8 horizontal glass plates ( $12 \times 14$  cm) which were then dried at 100° for 30 min. The thickness of the ayers was about 250  $\mu$ .